

# Validation of a RAPD marker linked to the anthracnose resistant gene *Co-5* in the common bean

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## ABSTRACT

Cultivar TU, one of the differential cultivars for anthracnose, carries the resistance gene *Co-5* that expresses resistance to a wide range of pathotypes in Brazil. The main goals of this study were to analyze the inheritance of resistance to anthracnose in the cultivar TU in segregating populations derived from crosses with cultivar Rudá, and to validate the random amplified polymorphic DNA (RAPD) marker OPB03<sub>450C</sub>, previously identified as being linked to resistance gene *Co-5* in cultivar Selection 1360. Results revealed that one single gene present in cultivar TU, *Co-5* is responsible for resistance, and that the band of 450 bp of primer OPB03 was linked to it at 15.4 cM. As OPB03<sub>450</sub> is not close enough from the *Co-5* gene of TU, it will not be useful during the pyramidation programs. However, this molecular marker can be useful for identification of the *Co-5* allele in other cultivars that carry this gene.

**KEY WORDS:** *Colletotrichum lindemuthianum*, *Phaseolus vulgaris* L., molecular markers, MAS, cultivar TU.

## INTRODUCTION

The common bean (*Phaseolus vulgaris* L.) is the main source of vegetable protein in most Latin American and African countries. The Brazilian population is the world largest consumer, with an annual average intake of 22-23 kg/person (Ventura and Costa, 1992; Borém and Carneiro, 1998). Fungal diseases are one of the main problems affecting common bean productivity, and anthracnose, caused by the fungus *Colletotrichum lindemuthianum* (Sacc. & Magn.) Scrib., is among the main diseases of the common bean in Brazil and in other growing regions of the world (Pastor-Corrales, 1985).

New breeding strategies, such as MAS (marker-aided selection) and pyramidation of resistance genes assisted by molecular markers, have been proposed as alternative solutions for development of varieties with long-lasting resistance (Haley et al., 1993; 1994; Michelmore, 1995; Johnson and Gepts, 1994; Kelly, 1995). In Brazil, the gene *Co-5* of cultivar TU has proven to be effective against 26 out of 27 *C. lindemuthianum* races identified (Rava et al., 1994; Thomazella et al., 2000). Common bean breeding programs aiming at pyramiding resistance genes could use this gene. Using allelism tests, it was possible to confirm allelic interrelations between gene *Co-5* from cultivar TU and one of the three resistance

genes present in cultivar G 2333, the one that is present in Selection 1360 (Young and Kelly, 1996). The cultivars TU and G 2333 are part of 12 international differentials common bean cultivars for anthracnose (Pastor-Corrales, 1992). The RAPD molecular marker OPB03<sub>450C</sub> was found to be linked to *Co-5* gene in a population derived from Selection 1360 (Young and Kelly, 1997). However, the distance between OPB03<sub>450C</sub> and *Co-5* in cultivar TU has not yet been determined.

The main goal of this study was to define the resistance inheritance pattern to *C. lindemuthianum* pathotype 89 in segregating populations derived from crosses between cultivar TU (resistant) and cultivar Rudá (susceptible to most *C. lindemuthianum* races) and to validate the RAPD marker OPB03<sub>450C</sub> previously identified as being linked to the resistance gene *Co-5* in cultivar Selection 1360.

## MATERIAL AND METHODS

### Source of *C. lindemuthianum* isolates and culture conditions

The isolate of pathotype 89 of *C. lindemuthianum* used in this work was collected in the Viçosa region (state of Minas Gerais, Brazil) and identified in our bean-breeding program (BIOAGRO/UFV). To

increase the amount of spores, the isolates were cultivated for approximately 10 days in a sterile medium containing common bean green pods. To confirm the identity of the isolate, it was inoculated in a 12-common-bean differential series according to Pastor-Corrales (1992).

### Common bean genetic material, crosses and seed production

Seeds from the differential cultivar TU and the susceptible cultivar Rudá were provided by CIAT (Tropical Agriculture International Center, Cali, Colombia) and EMBRAPA (Goiânia, GO, Brazil) respectively. Cultivar TU is a Mesoamerican, indeterminate prostrate with small black seeds, and Rudá is also a Mesoamerican, indeterminate commercial cultivar with "carioca type-seed". TU was used as male parent and crossed with Rudá under greenhouse conditions. All populations were kept in the greenhouse.

### Genetic analyses and evaluation of disease symptoms

The F<sub>2</sub> seeds of cross Rudá vs. TU and those of their respective parents were sown in the greenhouse. Primary leaves from each individual were collected, identified and stored at -80°C. Spores (1.2 x 10<sup>6</sup> conidia/ml) of pathotype 89 of *C. lindemuthianum* were sprayed with a De Vilbiss No. 15 apparatus to one primary leaf on 10-day-old plants. The plants were incubated in a mist chamber (20-22° C, 100% relative humidity) for seven days and then the disease symptoms were visually scored using a 1-to-9 scale. Resistant (R) phenotype was assigned to plants with no or limited symptoms (grades 1 to 3) whereas plants graded 4 or higher were considered

to be susceptible (S) (Rava et al., 1993).

### DNA extraction and amplification

DNA extraction was according to Doyle and Doyle (1990). Amplification reactions (Williams et al., 1990) were performed in a 9600 thermocycler (Perkin-Elmer, Norwalk, CT, USA) according to Alzate-Marin et al. (2001).

### Validation of RAPD markers

The Bulked Segregant Analysis technique (Michelmore et al., 1991) was used to validate the RAPD marker linked to resistance gene Co-5. Two contrasting bulks were prepared, each containing DNA from eight resistant or susceptible F<sub>2</sub> individuals. For DNA amplification, the primer OPB03 was used (Operon Technologies Inc., Alameda, CA, USA).

### Linkage analyses

Chi-square analyses were used to test the phenotypic segregation of the F<sub>2</sub> population originated from the cross Rudá/TU and to determine possible linkages between the RAPD marker OPB03<sub>450C</sub> and the resistance gene(s). Genetic distances between the markers and the resistance gene(s) were determined with the aid of the program MAP-MAKER III (Lander et al., 1987), using a LOD score minimum of 3.0.

## RESULTS AND DISCUSSION

The segregation analyses in the F<sub>2</sub> population derived from cross Rudá vs. TU revealed a 3:1 ratio

**Table 1.** Segregation analyses of RAPD marker OPB03<sub>450C</sub> and Co-5 gene in an F<sub>2</sub> population from the cross Rudá/TU inoculated with pathotype 89 of *C. lindemuthianum*.

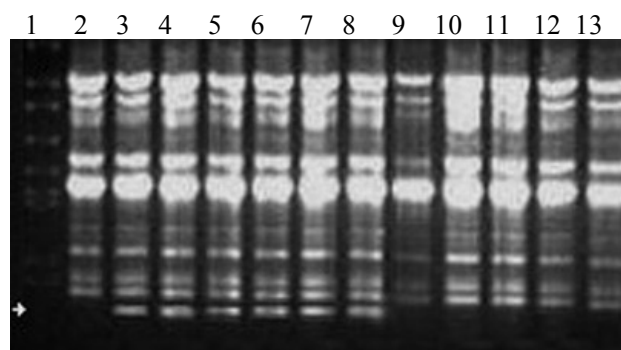
Locus Tested	Generation	Expected ratio	Observed ratio	$\chi^2$	P	cM <sup>1/</sup>
Co-5	F <sub>2</sub>	3:1	61:23	0.25	61.70	...
OPB03 <sub>450C</sub> <sup>2/</sup>	F <sub>2</sub>	3:1	60(+):24(-) <sup>3/</sup>	0.57	45.00	...
Co-5/OPB03 <sub>450C</sub> <sup>2/</sup>	F <sub>2</sub>	9:3:3:1 <sup>4/</sup>	55:6:5:18	45.59	0.00	15.4

<sup>1/</sup> Distance in centimorgans in relation to Co-5 (resistance gene); <sup>2/</sup> Molecular marker linked in coupling phase (c) to Co-5; <sup>3/</sup> (+) Band present; (-) Band absent; <sup>4/</sup> Ratio expected (9R+:3R-:3r+:1r-) if the resistance gene Co-5 and one marker (OPB03<sub>(450)</sub>) were independently inherited. The symbol + represents band present and the symbol -, band absent.

suggesting the involvement of one dominant gene (Co-5) determining resistance to pathotype 89 of *C. lindemuthianum* (Table 1). In resistant and susceptible bulks of this segregating population, one RAPD molecular marker of 450 bp linked in coupling (c) and generated by primer OPB03 was verified (Figure 1). Co-segregation analyses in the F<sub>2</sub> population revealed that marker OPB03<sub>450C</sub> was located 15.4 cM from the resistance gene (Table 1).

The RAPD molecular marker OPB03<sub>450C</sub>, linked to Co-5, was identified and validated in populations derived from cultivar Selection 1360 and BC<sub>1</sub>F<sub>2</sub> lines carrying the Co-5 gene derived from cultivar G 2333, respectively. In segregating populations derived from Selection 1360 in crosses with Black Magic and Blackhawk, distances of 7.2 and 4.4 cM were observed, respectively (Young and Kelly, 1997). However, in BC<sub>1</sub>F<sub>2</sub> lines derived from crosses between Rudá and G 2333, the distance was 16.4 cM (Alzate-Marín et al., 2001). These differences must have been caused by new recombination events when different female progenitors were used. In the present study, cultivar TU was used as an alternative resistance source, and a distance of 15.4 cM was observed. This distance was found to be similar to that observed in BC<sub>1</sub>F<sub>2</sub> lines derived from crosses between Rudá and G 2333.

As Co-5 has proven to be effective against a large number of Brazilian isolates of *C. lindemuthianum*, the validation of marker OPB03<sub>450C</sub> linked to the major gene Co-5 is a contribution for breeding programs which aim at pyramiding resistance genes



**Figure 1.** Electrophoresis analysis of amplification products obtained with primer OPB03. Lanes are as follows: 1, lambda DNA cut with EcoRI, BamHI and HindIII (size markers), 2, Rudá, 3, TU, 4-8, F<sub>2</sub> plants resistant to *C. lindemuthianum* pathotype 89, 9-13, F<sub>2</sub> plants susceptible to pathotype 89. The arrow indicates a DNA band of 450 bp linked in coupling phase to the resistance gene.

to anthracnose in common beans. However, as OPB03<sub>450</sub> is 15.4 cM distant from the Co-5 allele of TU, it will not be useful during the pyramiding of anthracnose resistance genes that make use of the progenitors analyzed in this study. However, this molecular marker can be useful for the identification of the Co-5 allele in other populations derived from crosses with G 2333, Selection 1360 and TU.

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## RESUMO

### Validação de um marcador RAPD ligado ao gene Co-5 de resistência à antracnose do feijoeiro comum

O cultivar TU, um dos cultivares diferenciadores para antracnose do feijoeiro, possui o gene Co-5 o qual confere resistência a um amplo número de patótipos no Brasil. Os principais objetivos deste trabalho foram estudar a herança da resistência a antracnose do cultivar TU em populações segregantes derivadas do cruzamento com o cultivar Rudá, e validar o marcador molecular RAPD OPB03<sub>450C</sub>, previamente identificado como ligado ao gene Co-5 no cultivar Seleção 1360. Os resultados revelaram que um único gene do cultivar TU, o Co-5, é responsável pela resistência, e que a banda de 450 pb do primer OPB03 está ligada ao gene a uma distância de 15,4 cM. Como OPB03<sub>450C</sub> está distante do gene Co-5 do cultivar TU, não poderá ser utilizado em programas de piramidação de genes. No entanto, este marcador molecular poderá ser utilizado na identificação do alelo Co-5 em outros cultivares que possuam este gene.

## REFERENCES

Alzate-Marín, A.L.; Menarim, H.; Baía, G.S.; Paula Jr., T.J.; Souza, K.A. De; Costa, M.R.; Barros, E.G. and Moreira, M.A. 2001. Inheritance of anthracnose resistance in the common bean differential cultivar G 2333 and identification of a new molecular marker linked to the Co-4<sup>2</sup> gene. *Journal of Phytopathology*. 149:259-264.

- Borém, A. and Carneiro, J.E.S. 1998. A Cultura. p.13-17. In: Vieira, C.; Paula-Jr, T. J. and Borém, A. (Eds). Feijão: aspectos gerais e cultura no estado de Minas. Ed. UFV, Viçosa.
- Doyle, J.J. and Doyle, J.L. 1990. Isolation of plant DNA from fresh tissue. *Focus*. 12:13-15.
- Haley, S.D.; Miklas, P.N.; Stavely, J.R.; Byrum, J. and Kelly, J.D. 1993. Identification de RAPD markers linked to a major rust resistance gene block in common bean. *Theoretical and Applied Genetics*. 86:505-512.
- Haley, S.D.; Afanador, L.K. and Kelly, J.D. 1994. Selection for monogenic resistance traits with coupling and repulsion-phase RAPD markers. *Crop Science*. 34:1061-1066.
- Johnson, W.C. and Gepts, P. 1994. Two new molecular markers linked to bc-3. *Annual Report Bean Improvement Cooperative -BIC*. 37:206-207.
- Kelly, J.D. 1995. Use of random amplified polymorphic DNA markers in breeding for major resistance to plant pathogens. *HortScience*. 30:461-465.
- Lander, E.S.; Green, P.; Abrahamson, J.; Barlow, A.; Daly, M.J.; Lincon, S.E. and Newburgh, L. 1987. MAPMAKER: An interactive computer package for constructing primary genetic linkage maps of experimental and natural populations. *Genomics*. 1:174-181.
- Michelmore, R. 1995. Molecular approaches to manipulation of disease resistance genes. *Annual Review of Phytopathology*. 15:393-427.
- Michelmore, R.; Paran, I. and Keselli, V. 1991. Identification of markers linked to disease-resistance genes by bulked segregant analysis: A rapid method to detect markers in specific genomic regions by using segregating populations. *Proceedings of the National Academy of Sciences of the United States of America*. 88:9828-9832.
- Pastor-Corrales, M.A. 1985. Enfermedades del frijol causadas por hongos. p.172-180. In: López, M.; Fernández, F. and Schoonhoven A. (Eds.). Frijol: Investigación y Producción. PNUD-CIAT, Cali.
- Pastor-Corrales, M.A. 1992. Recomendaciones y acuerdos del primer taller de antracnosis en América Latina. p.240-250. In: Pastor-Corrales, M. (Ed.). La antracnosis del frijol común, *Phaseolus vulgaris*, en América Latina. Doc. de trabajo, 113. CIAT, Cali.
- Rava, C.A.; Molina, J.; Kauffmann, M. and Briones, I. 1993. Determinación de raças fisiológicas de *Colletotrichum lindemuthianum* en Nicaragua. *Fitopatologia Brasileira*.18:388-391.
- Rava, C.; Purchio, A. and Sartorato, A. 1994. Caracterização de patótipos de *Colletotrichum lindemuthianum* que ocorrem em algumas regiões produtoras de feijoeiro comum. *Fitopatologia Brasileira*.19:167-172.
- Thomazella, C.; Gonçalves-Vidigal, M.C.; Vida, J. B.; Vidigal Filho, P.S. and Rimoldi, F. 2000. Identification of *Colletotrichum lindemuthianum* races in *Phaseolus vulgaris* L. *Annual Report Bean Improvement Cooperative -BIC*. 43:82-83.
- Ventura, J. and Costa, H. 1992. Situação atual do feijoeiro e da antracnose no Estado do Espírito Santo, Brasil. p.69-85. In: Pastor-Corrales, M. (Ed.) La antracnosis del frijol común, *Phaseolus vulgaris*, en América Latina. Doc. de trabajo, 113. CIAT, Cali.
- Williams, J.; Kubelik, A.; Livak, K.; Rafalski, A. and Tingey, S. 1990. DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. *Nucleic Acids Research*. 18:6531-6535.
- Young, R. and Kelly, J.D. 1996. Characterization of the genetic resistance to *Colletotrichum lindemuthianum* in common bean differential cultivars. *Plant Disease*. 80:650-654.
- Young, R. and Kelly, J.D. 1997. RAPD markers linked to three major anthracnose resistance genes in common bean. *Crop Science*.37:940-946.

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